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UV-B-Induced Inflammasome Activation Can Be Prevented by Cis-Urocanic Acid in Human Corneal Epithelial Cells

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PURPOSE. The cornea is continually exposed to highly energetic solar UV-B (280–320 nm). Our aim was to investigate whether UV-B triggers the activation of NLRP3 inflammasomes and the production of IL-1 β and/or IL-18 in human corneal epithelial (HCE) cells. Additionally, we studied the capability of cis-urocanic acid (cis-UCA) to prevent inflammasome activation or alleviate inflammation through other signaling pathways.

METHODS. HCE-2 cell line and primary HCE cells were primed using lipopolysaccharide or TNF- α . Thereafter, cells were exposed to UV-B before or after the addition of cis-UCA or caspase-1 inhibitor. Caspase-1 activity was measured from cell lysates by an enzymatic assay. IL-1 β , IL-18, IL-6, IL-8, and NLRP3 levels were detected using the ELISA method from cell culture media. Additionally, intracellular NLRP3 levels were determined by the Western blot technique, and cytotoxicity was measured by the LDH assay.

RESULTS. UV-B exposure significantly increased caspase-1 activity in TNF- α -primed HCE cells. This result was consistent with the concurrently induced IL-1 β secretion. Both caspase-1 activity and release of IL-1 β were reduced by cis-UCA. Additionally, UV-B stimulated the caspase-1-independent production of IL-18, an effect also reduced by cis-UCA. Cis-UCA decreased the release of IL-6, IL-8, and LDH in a time-dependent manner when administered to HCE-2 cells after UV-B exposure.

CONCLUSIONS. Our findings demonstrate that UV-B activates inflammasomes in HCE cells. Cis-UCA can prevent the secretion of IL-1 β and IL-18 and therapeutically reduces the levels of IL-6, IL-8, and LDH in UV-B-stressed HCE cells.

Keywords: ultraviolet B, inflammasome, urocanic acid, cornea, inflammation

The cornea absorbs a significant amount of ambient UV-B (290–320 nm) and protects the deeper ocular structures from harmful radiation.^{1–3} Although UV-B accounts for only 4% of terrestrial light, it is light with a highly energetic wavelength, causing many harmful eye conditions, such as painful photokeratitis and climatic droplet keratopathy.³⁴ At the cellular level, excessive UV-B evokes corneal phototoxicity including direct DNA damage, such as the formation of cyclobutane pyrimidine dimers⁵ and disturbances in the antioxidant balance resulting in cellular oxidative stress.^{6–8}

IL-1 β and IL-18 are pro-inflammatory cytokines that have a major role in inflammatory responses.^{9,10} The secretion of these cytokines is regulated by intracellular multiprotein platforms called inflammasomes. Nucleotide-binding domain and leucine-rich repeat pyrin-containing protein 3 (NLRP3) is the receptor of the most extensively studied inflammasome^{11,12}; it can be activated in response to nonsterile (microbes) or sterile stressors.¹¹ NLRP3 activa-

tion has to be preceded by a priming step,^{11,12} which triggers the translation of NLRP3 and pro-IL-1 β via nuclear factor-kappa B (NF- κ B) signaling.¹³ IL-18 precursor protein is constitutively expressed in most of human cells (e.g., in the corneal epithelial cells, peripheral blood mononuclear cells, keratinocytes, endothelial cells, and retinal pigment epithelial cells).^{14–17}

Inflammasome activation occurs when primed cells sense pathogen-associated molecular patterns or danger-associated molecular patterns via the NLRP3, which leads to NLRP3 oligomerization and the assembly of the inflammasome multiprotein complex also containing the adaptor protein apoptosis-associated speck-like protein containing a C-terminal caspase recruitment domain (ASC) and the cysteine protease procaspase-1.^{18,19} The formation of the inflammasome complex causes the proteolytic activation of caspase-1,^{20,21} which enables the cleavage of pro-IL-1 β into its biologically active and secreted form.²¹ In conjunction

with the cleavage of pro-IL-1 β , pro-IL-18 requires proteolytic processing before secretion.²²

Urocanic acid (UCA) is a naturally occurring endogenous chromophore produced from histidine in the upper layer of the epidermis in a deamination reaction catalyzed by histidase.²³ UCA usually occurs as a *trans* form, but is converted to the *cis* isomer in a photoisomerization reaction by UV-B.^{23,24} Therapeutic effects of *cis*-UCA have been studied for nearly three decades. Despite the fact that the effects of *cis*-UCA may be cell type-dependent, many *in vivo* and *in vitro* studies have revealed its anti-inflammatory properties and potential to protect from cell injury.^{25–28} Our previous studies have revealed that *cis*-UCA is well tolerated by human corneal epithelial (HCE) and conjunctival epithelial (HCEC) cells.²⁹ Additionally, pretreatment with *cis*-UCA prevents cell death and the secretion of IL-8 and IL-6 in UV-B-induced HCE and HCEC cells.²⁹ Moreover, we have shown that *cis*-UCA could reduce the activation of activator protein-1 and mitogen-activated protein kinase pathways in the UV-B-irradiated HCE-2 cell line.³⁰

Although it is evident that excessive UV-B exposure can induce an acute inflammatory response in the cornea, its role in inflammasome signaling is unknown. Because inflammasomes are key players in inflammation, we have now investigated whether the UV-B-induced inflammation is regulated by the inflammasomes in HCE cells and whether it can be prevented by *cis*-UCA. We have also explored the therapeutic potential of *cis*-UCA by investigating whether it reduces IL-6, IL-8, or LDH release when administered after UV-B exposure.

MATERIALS AND METHODS

Cell Stimulations

The human corneal epithelial cell line (HCE-2) was purchased from the American Type Culture Collection. HCE-2 cells were cultured in the Keratinocyte Serum Free growth medium (Life Technologies, Paisley, UK) containing 50 μ g/mL bovine pituitary extract, 5 ng/mL human recombinant epidermal growth factor 1-53 (EGF 1-53; both from Life Technologies, Grand Island, NY, USA), 100 U/mL penicillin (Lonza, Walkersville, MD, USA), 100 μ g/mL streptomycin (Lonza), and 0.005 mg/mL insulin (Sigma Aldrich, Saint Louis, MO, USA) at 37°C in a humidified atmosphere with 5% CO₂. Cells in passage numbers ranging from 69 to 86 were used in the study. HCE cell culture plates for the maintenance (100 mm x 20 mm; Sigma Aldrich, St. Louis, MO, USA) and 12-well culture plates for the experiments (Corning Inc., Corning, NY, USA) were coated with 0.01 mg/mL fibronectin (Sigma-Aldrich), 0.03 mg/mL collagen (STEMCELL technologies, Vancouver, Canada), and 0.01 mg/mL bovine serum albumin (Roche Diagnostics GmbH, Mannheim, Germany) in the Keratinocyte Serum Free Medium. The coating solution was incubated for 30 to 90 minutes at 37°C until replaced by cell suspensions. In the experiments, cells were seeded on 12-well plates at a density of 1.5×10^5 cells/mL and incubated for 24 hours in a humidified 5% CO₂ incubator at 37°C.

All tissue collections complied with the Guidelines of the Helsinki Declaration and were approved by the Local Ethical Committees (No 2017/418). Human limbal biopsies, obtained from cadaveric corneo-scleral rings after corneal transplantation were treated with Dispase II (2.4 U/mL, Roche Diagnostics) for 10 minutes at 37°C and thereafter blocked with fetal bovine serum (Sigma-Aldrich), plated on

six-well plates (Corning Inc.) with the epithelial side down and covered with cell culture medium. Once the limbal biopsies were attached, they were completely covered and maintained in DMEM/F12 medium (Thermo Fisher Scientific, Waltham, MA, USA) containing 100 U/mL penicillin, 100 μ g/mL streptomycin, 1.25 μ g/mL amphotericin B, 5% FBS, 2 ng/mL human epidermal growth factor, 5 μ g/mL insulin-transferrin-sodium selenite, 15 μ M hydrocortisone, 0.5% dimethylsulfoxide, and 30 ng/mL cholera toxin A (all from Sigma Aldrich) in a humidified 5% CO₂ incubator at 37°C. After the primary HCE (pHCE) cells growing out from the limbal biopsies had reached confluency, they were treated with 0.25% Trypsin-EDTA (Sigma Aldrich) and seeded on 12-well plates (Corning Inc.) at a density of 2.5×10^5 cells/mL and incubated for 48 to 72 hours in a humidified 5% CO₂ incubator at 37°C. Thereafter, the cell layers were washed and replaced with DMEM/F12 medium (Thermo) without any supplements.

The priming signal for pHCE and HCE-2 cells was provided by tumor necrosis factor α (TNF- α , 10 ng/mL; R&D Systems, Minneapolis, MN, USA) or lipopolysaccharide (LPS, 1 μ g/mL; Sigma Aldrich) by incubating cells for 24 h in a humidified 5% CO₂ incubator at 37°C. Thereafter, cell cultures were irradiated with UV-B (two TL 20W/12 tubes, Philips, Eindhoven, The Netherlands) at the energy level of 0.2 J/cm². The UV-B lamps were calibrated using a spectroradiometer (Waldman Variocontrol, Germany) by Istekki Ltd. (Kuopio, Finland) before the experiments. The irreversible caspase-1 inhibitor, Ac-YVAD-CMK (200 μ M for HCE-2 cells; Calbiochem, San Diego, CA, USA; or 100 μ M for pHCE cells; Sigma Aldrich) was added to the wells 1 hour before the UV-B irradiation, where applicable. Additionally, cells were treated with *cis*-urocanic acid (*cis*-UCA, 100 μ g/mL, Sigma Aldrich) 1 hour before or immediately after the UV-B exposure. The concentration of *cis*-UCA had been optimized in our previous study.²⁹

Sample Collection

Cell culture medium samples and whole cell lysates were collected 0.5 to 48 hours after the exposure to UV-B or *cis*-UCA, depending on the experiment. Cell culture medium samples were centrifuged at $380 \times g$ for 10 minutes and supernatants were transferred into clean tubes. The cells were rinsed with ice-cold 1x Dulbecco's phosphate-buffered saline (Sigma Aldrich) and lysed by addition of the M-PER solution (Thermo Fisher Scientific) or the cell lysing buffer provided by the manufacturer of the caspase-1 activity kit (R&D). Cell lysates prepared using the M-PER solution were centrifuged at $16\,000 \times g$ for 10 minutes at 4°C. In the measurements of caspase-1 activity, cells from two to six nonirradiated or four to 12 UV-B-irradiated parallel wells were collected into one tube and centrifuged according to the manufacturers protocol. Before the analyses, protein concentrations were measured using the Bradford method.³¹

ELISA Measurements

IL-1 β , IL-6, IL-8 (BD Biosciences, San Diego, CA, USA), IL-18 (eBioscience, San Diego, CA, USA), and NLRP3 (Cusabio, Wuhan, China) were measured using the ELISA method from the cell culture medium according to the manufacturer's instructions. Absorbance values were measured at 450 nm with the reference wavelength of 655 nm by a spectrophotometer (Biorad, Model 550, Hercules, CA, USA).

Caspase-1 Activity

Caspase-1 activity was measured using the caspase-1 colorimetric assay according to the manufacturer's protocol. Absorbance values were determined at a wavelength of 405 nm by a spectrophotometer (ELx808, Biotek Instruments Inc., Winooski, VT, USA). The kit supplements without cell lysates were used as blank samples. Thereafter, blank-subtracted results were normalized to protein concentrations and expressed as a ratio to the untreated cells.

Cytotoxicity Measurements

The quantity of LDH was measured from cell culture medium samples using the CytoTox96 Non-Radioactive Cytotoxicity assay (Promega Corporation, Madison, WI, USA) according to the manufacturer's instructions. Colorimetric reactions were monitored at a wavelength of 490 nm in a spectrophotometer (Biorad).

Western Blot

Equal amounts of protein (20–40 µg in HCE-2 cell lysates or 5–10 µg in primary HCE cell lysates) were run in 10% SDS-PAGE gels and wet-blotted overnight onto nitrocellulose membranes (GE Healthcare, Little Chalfont, Buckinghamshire, UK). Ponceau S (Sigma-Aldrich) staining was used to ensure the accurate protein transfer from the gels to the membranes. The membranes were blocked with 5% milk in 0.1% Tween-20/Tris-buffered saline for 1 hour at room temperature (RT). The Tween/Tris-buffered saline solution was also used as a washing buffer and a diluent for the antibodies. Thereafter, the membranes were incubated with an anti-CIAS1/NALP3 (1:1000; cat. 109314, Abcam, Cambridge, UK) monoclonal rabbit antibody overnight at +4°C. The membranes were washed 3 × 5 minutes with washing buffer followed by a 2-hour incubation at RT with a horseradish peroxidase-conjugated anti-rabbit IgG antibody (1:5 000, cat. A16104, Novex) in blocking buffer. The washing steps were repeated and the NLRP3 protein-antibody complexes were detected using the enhanced chemiluminescent assay after the addition of the substrate (Millipore, Billerica, MA, USA). The band intensities given by NLRP3 were normalized to α -tubulin values using a monoclonal mouse α -tubulin (1:8000; cat. T5168, Sigma Aldrich). Following the incubation for 15 minutes at RT, membranes were washed three times for 5 minutes. Polyclonal horseradish peroxidase-conjugated sheep anti-mouse IgG (1:10 000; cat. NA931, GE Healthcare) was used as a secondary antibody and incubated for 15 minutes at RT before performing the washing and detection steps as described previously. All the results were quantified using the ImageJ program (<http://rsb.info.nih.gov/ij/index.html>).

Confocal Imaging of DsRed-ASC

HCE-2 cells were cultured on eight-well chamber slides (Ibidi GmbH, Martinsried, Germany) at a density of 4.5×10^4 cells/well. Sub-confluent (70%–80%) cell layers were transfected with DsRed-ASC plasmid construct (20 ng) along with ExGen500 transfection reagent (Thermo Fischer Scientific, Waltham, MA, USA). The cells were incubated for 24 hours in a humidified 5% CO₂ incubator at 37°C followed by a priming with TNF- α (10 ng/mL) for the next 24 hours. Thereafter, ASC specks were observed under the confocal microscope

(Zeiss Axio Observer inverted microscope with Zeiss LSM 700 confocal module) at 5 and 24 hours upon UV-B exposure (0.2 J/cm²).

Statistical Analyses

All results were analyzed using the GraphPad Prism program (GraphPad Software Version 7.04, San Diego, CA, USA). Pairwise comparisons were performed using the Mann-Whitney *U*-test, and *P* values ≤ 0.05 were considered statistically significant.

RESULTS

UV-B Exposure Induces Caspase-1 Dependent Secretion of IL-1 β in HCE Cells

To examine the ability of UV-B to activate inflammasomes in HCE-2 cells, cell cultures were primed with LPS or TNF- α and subsequently exposed to UV-B light. LPS and TNF- α -primed and UV-B-treated HCE-2 cells secreted higher amounts of IL-1 β (1.6-fold in the LPS-treated cells and 3.0-fold in the TNF- α -primed cells) compared with LPS or TNF- α -treated controls (mean for LPS: 0.3 pg/mL vs. LPS + UV-B: 0.4 pg/mL, *P* < 0.001, Fig. 1 A; TNF- α : 0.5 pg/mL vs. TNF- α + UV-B: 1.7 pg/mL, *P* < 0.0001, Fig. 1 B). Additionally, UV-B without priming tripled the IL-1 β levels when compared to untreated control cells (mean for untreated control: 0.1 pg/mL vs. UV-B: 0.3 pg/mL; Figs. 1 A–B) although the levels were 1.4 (*P* < 0.01) and 4.1 (*P* < 0.0001) times lower when compared to LPS or TNF- α -primed and UV-B-treated cells, respectively.

The results with the pHCEs were in line with those obtained with HCE-2 cells, showing that the secretion of IL-1 β was increased after the UV-B exposure (untreated control: 22.2 pg/mL vs. UV-B: 111.0 pg/mL, *P* < 0.0001; TNF- α : 52.0 pg/mL vs. TNF- α + UV-B: 326.2 pg/mL, *P* < 0.0001; Fig. 1 C). Once again, TNF- α + UV-B-treated cells significantly augmented the production of IL-1 β when compared with UV-B-treated cells without priming (*P* < 0.0001; Fig. 1 C). Overall, TNF- α + UV-B-treatment induced a much greater, (i.e., 190-fold elevations) response of IL-1 β in pHCE cells (TNF- α + UV-B: 326.2 pg/mL) in comparison to the HCE-2 cell line (TNF- α + UV-B: 1.7 pg/mL). LDH data showed that the increased cytokine levels were not attributable to the passive leakage of proteins (UV-B vs. TNF- α + UV-B, *P* > 0.05; Fig. 1 D). Moreover, the addition of a selective caspase-1 inhibitor significantly mitigated the secretion of IL-1 β in UV-B-irradiated HCE-2 (*P* < 0.001, Fig. 1 E) and cultured primary HCE cells (*P* < 0.05; Fig. 1 F), evidence of a role in inflammasome signaling in the production of IL-1 β . It was confirmed with the LDH assay that caspase-1 inhibitor was not toxic to the HCE cells (TNF- α + UV-B vs. TNF- α + caspase-1 inhibitor + UV-B, *P* > 0.05; Fig. 1 G).

UV-B Induces the Caspase-1-Independent Secretion of IL-18 in Human Corneal Epithelial Cells

Next, we evaluated whether the secretion of IL-18 was upregulated after the UV-B exposure in HCE cells. Instead of LPS priming, only TNF- α was used to prime cells since it resulted in 4.25 times higher concentration of IL-1 β in

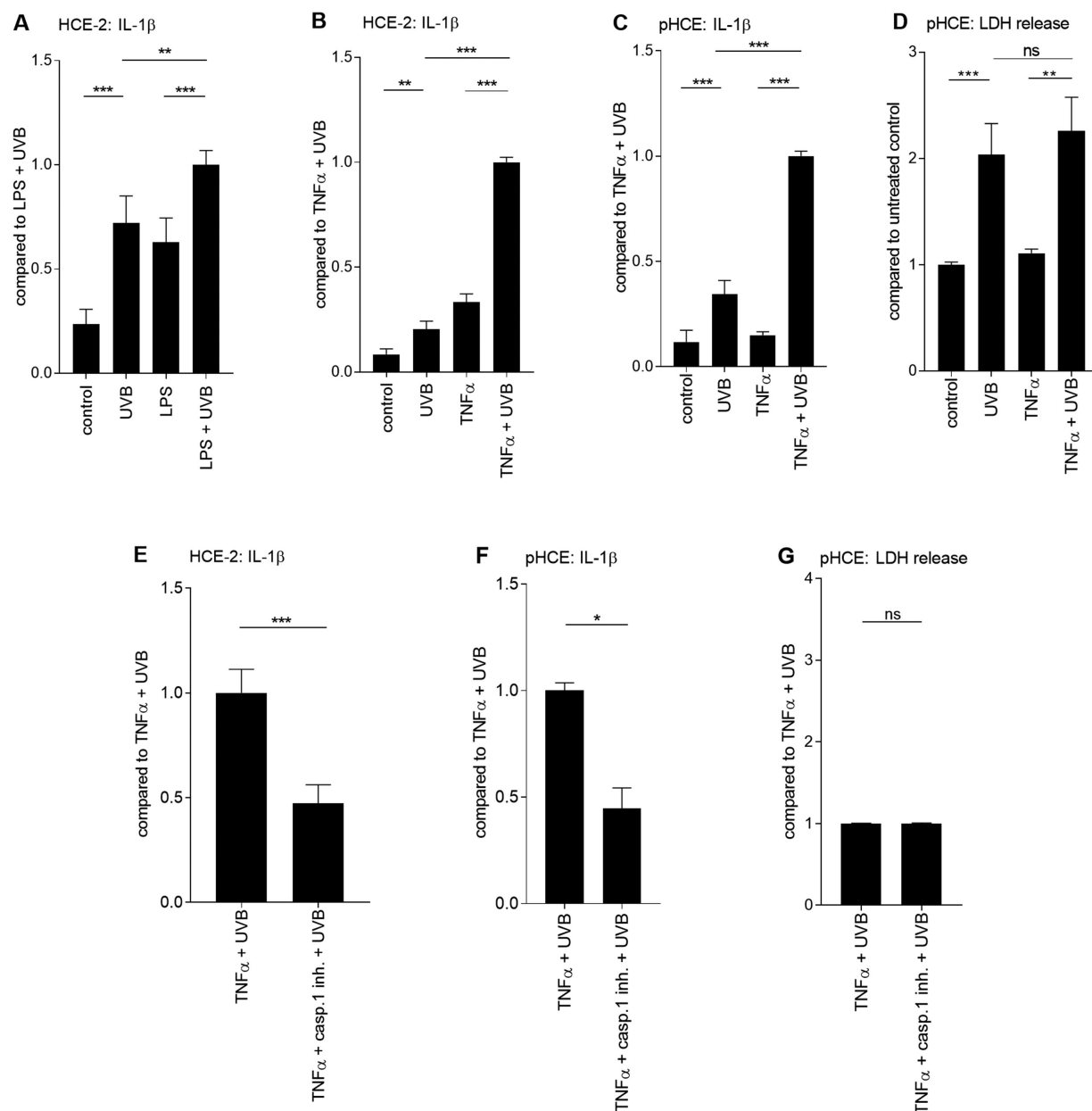


FIGURE 1. Caspase-1-dependent secretion of IL-1β in UV-B-stimulated HCE-2 and primary HCE cells. HCE-2 cells were primed for 24 hours either with LPS (1 μg/mL; A) or TNF-α (10 ng/mL; B) followed by irradiation with UV-B (0.2 J/cm²). The secretion of IL-1β was measured by the ELISA method from HCE cell culture media for 24 hours after the UV-B irradiation. The levels of IL-1β were combined from three independent experiments with five to six parallel samples per group. Additionally, levels of IL-1β and LDH were measured from the primary HCE cell culture media from eight donors with one to three parallel samples per donor (C-D). Alternatively, HCE-2 (E) or primary HCE (F-G) cells were pretreated with the caspase-1 inhibitor (200 μM [E] or 100 μM [F-G] for 1 hour before the UV-B exposure). IL-1β data from HCE-2 cells (E) have been combined from three independent experiments each with 10 to 12 parallel samples. IL-1β and LDH data from cultured primary HCE cells (F-G) have been combined from four donors with one to two parallel samples per donor. All data are presented as mean ± SEM. **P* < 0.05, ***P* < 0.01, ****P* < 0.001, ns = not significant, Mann-Whitney *U*-test; pHCE = primary HCE cells; casp.1 inh. = caspase-1 inhibitor.

UV-B-irradiated HCE-2 cells (LPS + UV-B: 0.4 pg/mL and TNF-α + UV-B: 1.7 pg/mL; **Figs. 1 A-B**). In line with the IL-1β findings, the secretion of IL-18 was increased after the UV-B irradiation (untreated control: 23.7 pg/mL, UV-B: 98.1 pg/mL; *P* < 0.01). Similarly, UV-B-induced the secretion of IL-18 in TNF-α-treated pHCE cells (TNF-α: 40.0 pg/mL, TNF-α + UV-B: 105.3 pg/mL; *P* < 0.001; **Fig. 2 A**). There was

no difference in the levels of IL-18 when UV-B and TNF-α + UV-B-treated cells were compared with each other (*P* > 0.05; **Fig. 2 A**). Surprisingly, treatment with the caspase-1 inhibitor did not reduce the secretion of IL-18 (*P* > 0.05; **Fig. 2 B**), suggesting that UV-B does not cause the maturation IL-18 via a caspase-1-dependent mechanism in HCE cells.

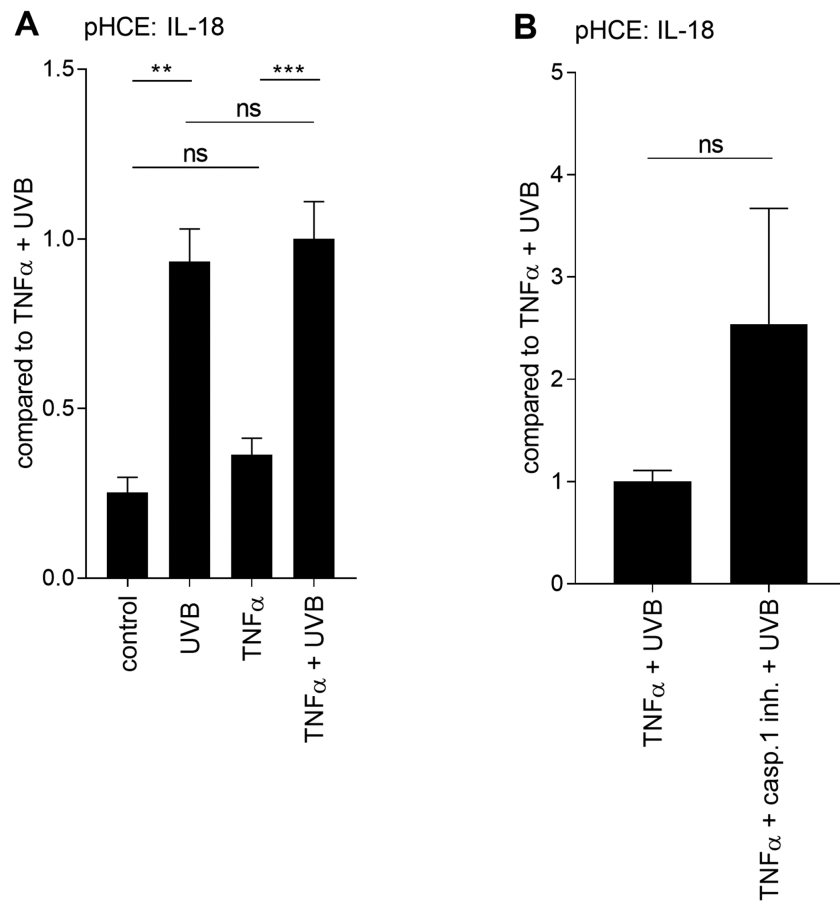


FIGURE 2. Caspase-1-independent release of IL-18 in UV-B-irradiated primary HCE cells. Primary HCE cells were treated with TNF- α (10 ng/mL) for 24 hours followed by irradiation under UV-B light (0.2 J/cm²). The secretion of IL-18 was measured for 24 hours after the UV-B exposure from the cell culture medium samples (A). Alternatively, cells were pretreated with the caspase-1 inhibitor (100 μ M) at 1 hour before UV-B irradiation (B). ELISA results of extracellular IL-18 have been combined from (A) three or (B) two donors with two parallel samples per donor. Data are represented as mean \pm SEM. ** P < 0.01, *** P < 0.001, ns = not significant, Mann-Whitney U -test; casp.1 inh. = caspase-1 inhibitor.

NLRP3 is Secreted After the UV-B Irradiation from TNF- α -Primed Human Corneal Epithelial Cells

TNF- α significantly increased the levels of intracellular NLRP3 in HCE-2 cells (P < 0.001; Fig. 3 A), suggesting an efficient priming signal for the cells. NLRP3 was released out of the cells following the inflammasome activation, as shown by the 2.3-fold reduction in the levels of intracellular NLRP3 (P < 0.01) and the 3.5-fold increase in the cell culture medium (P < 0.001) in TNF- α + UV-B-treated HCE-2 cells in comparison to TNF- α -treated control cells (Figs. 3 A-B). Additionally, TNF- α induced the secretion of NLRP3 in comparison to untreated controls (P < 0.001; Fig. 3 B), though the levels were 3.5 times lower when compared with the TNF- α + UV-B group. Exposure to UV-B without the priming signal was not essential for the induction of NLRP3 in HCE-2 cells since the intracellular or extracellular levels of NLRP3 showed no change upon exposure (P > 0.05, Figs. 3 A-B). Unlike HCE-2 cells, intracellular NLRP3 levels were increased in TNF- α -primed pHCE cells after the UV-B exposure (P < 0.05; Fig. 3 C). Moreover, NLRP3 was released out of the primed pHCE cells after the UV-B exposure, while instead of UV-B (P > 0.05; Fig. 3 D), TNF- α highly induced the secretion of NLRP3 (P < 0.001; Fig. 3 D) when compared with untreated control.

UV-B Affects Formation of ASC Specks in Human Corneal Epithelial Cells

Thereafter, we studied further the activation of NLRP3 by measuring ASC specks upon UV-B irradiation in HCE-2 cells under the confocal microscope. UV-B clearly induced the formation of ASC specks already 5 hours upon UV-B exposure when compared with untreated control (Figs. 4 A-C). Additionally, ASC specks were observed at the 24 h time-point upon UV-B irradiation in TNF- α + UV-B-irradiated cells but not in untreated control (Figs. 4 D-E). These findings support our other data that UV-B exposure in primed HCE cells activates the assembly of inflammasomes.

cis-UCA Prevents the UV-B-Induced Secretion of IL-1 β , IL-18, and LDH from Human Corneal Epithelial Cells

Next, the ability of cis-UCA to modulate the UV-B-induced inflammasome activation was studied. Cis-UCA reduced the UV-B-induced caspase-1 activity in HCE-2 cells (P < 0.05; Fig. 5 A). Subsequent experiments with the HCE-2 cell line revealed that cis-UCA reduced the production of IL-1 β (P < 0.001; Fig. 5 B). In an attempt to confirm these

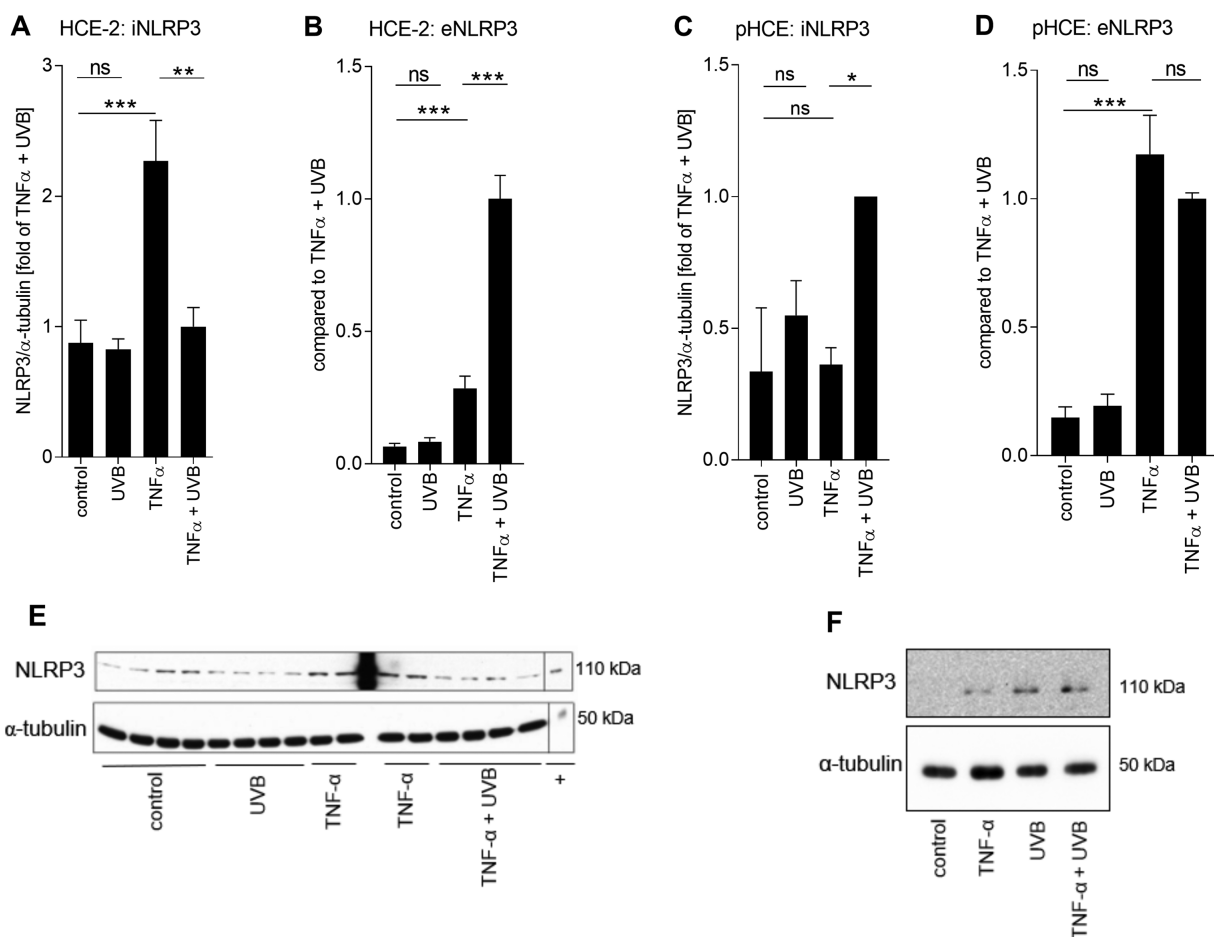


FIGURE 3. Fate of NLRP3 in UV-B-stressed HCE cells. HCE-2 cells were primed for 24 hours with TNF- α (10 ng/mL) following irradiation with UV-B (0.2 J/cm²), and then incubated for 24 hours before sample collection and analysis. Intracellular levels of NLRP3 (iNLRP3; A) were measured by the Western blot technique from cell lysates and the levels of NLRP3 were normalized to those of α -tubulin. Data in panel A is combined from two (UV-B) to four (untreated control, TNF- α , TNF- α + UV-B the correct group is TNF- α + UVB. This means that extra red comma should be removed.) independent experiments with three to four parallel samples in each group. Extracellular levels of NLRP3 (eNLRP3; B) were measured by the ELISA method from HCE-2 cell culture supernatants and data were combined from three independent experiments with three parallel samples in each group. Experiments were repeated using primary HCE cells, and intracellular levels of NLRP3 were measured by Western blot from cells of four donors (C), and extracellular levels of NLRP3 by ELISA from four donors with two parallel samples (D). (E, F) Representative Western blots from NLRP3 and α -tubulin measurements of (A) HCE-2 cells lysates or (C) primary HCE cell lysates. All data are presented as mean \pm SEM. * P < 0.05, ** P < 0.01, *** P < 0.001, ns = not significant, Mann-Whitney U -test; + the NLRP3 positive control + equals to the NLRP3 positive control.

findings, we found that cis-UCA decreased by 6.9 times the secretion of IL-1 β when compared with UV-B-treated pHCE cells without cis-UCA pretreatment (mean for TNF- α + UV-B: 203.1 pg/mL vs. TNF- α + cis-UCA + UV-B: 29.4 pg/mL, P < 0.001; Fig. 5 C). Actually, cis-UCA restored the release of IL-1 β back to the baseline level of non-irradiated pHCE cells (TNF- α : 26.5 pg/mL vs. TNF- α + cis-UCA + UV-B: 29.4 pg/mL, P > 0.05; Fig. 5 C).

In line with the IL-1 β results, cis-UCA reduced the IL-18 secretion by 3.9 times when compared with TNF- α + UV-B-treated cells without cis-UCA (TNF- α + UV-B: 91.5 pg/mL and TNF- α + cis-UCA + UV-B: 23.6 pg/mL, P < 0.05; Fig. 5 D). Moreover, cis-UCA restored the production of IL-18 back to the nonirradiated control level (P > 0.05; Fig. 5 D). Collectively, these results suggest that cis-UCA was able to inhibit the inflammasome-dependent secretion of IL-1 β as well as the inflammasome-independent secretion of IL-18.

In cell viability assays, cis-UCA significantly protected pHCE cells from UV-B-induced cell death (TNF- α + UV-B vs. TNF- α + cis-UCA + UV-B, P < 0.001; Fig. 5 E). In line with the cytokine results, LDH release remained at the baseline level if cis-UCA had been provided to the UV-B-irradiated pHCE cells (TNF- α vs. TNF- α + cis-UCA + UV-B, P > 0.05; Fig. 5 E).

cis-UCA has a Therapeutic Effect on the Release of IL-8, IL-6, and LDH in HCE-2 Cells when Applied After the UV-B Exposure

Finally, we investigated the therapeutic potential of cis-UCA by applying it to HCE-2 cell cultures after the UV-B irradiation. Cis-UCA significantly reduced the secretion of both IL-8 (P < 0.001; Fig. 6 A) and IL-6 (P < 0.01-0.001; Fig. 6 B) at all five time points (0.5, 4, 6, 24, and 48 hours) when

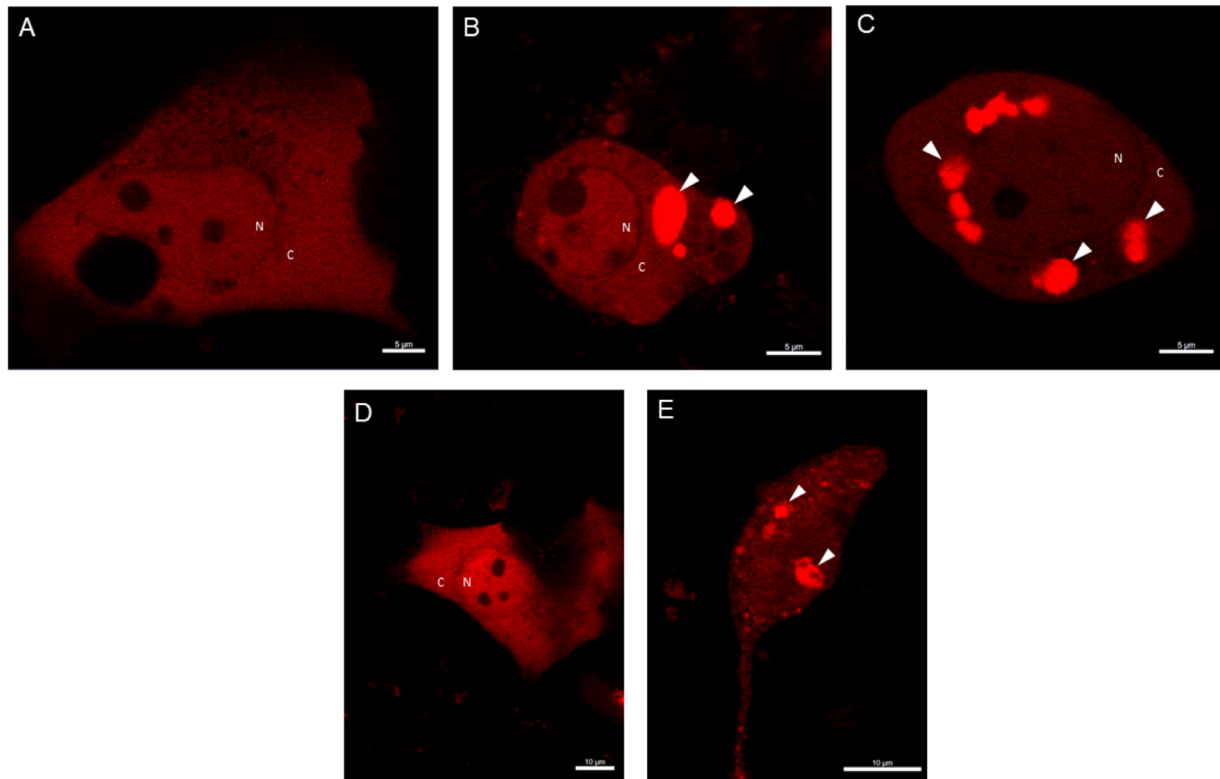


FIGURE 4. The formation of ASC specks upon UV-B irradiation in HCE cells. HCE-2 cells were transfected with DsRed-ASC plasmid constructs for 24 hours and primed for the next 24 hours with TNF- α (10 ng/mL). HCE-2 cells were observed under the confocal microscope 5 hours after the UV-B exposure (0.2 J/cm²; A = control group and B-C = TNF- α + UV-B treated group). Alternatively, microscopic examinations were performed 24 hours after the UV-B irradiation (D = control group and E = TNF- α + UV-B treated group). Experiments were repeated three times and images were photographed using the 63-fold objective (A-C: scale bar, 5 μ m; D-E: scale bar, 10 μ m). White arrows indicate ASC specks. N = nucleus, C = cytoplasm.

compared with cells exposed only to UV-B. Cis-UCA reduced the secretion of IL-8 and IL-6 even under the basal control level at the 0.5 hours time point (IL-8 from untreated control: 1 vs. UV-B + cis-UCA: 0.2, Fig. 6 A; IL-6 from untreated control: 1 vs. UV-B + cis-UCA: 0.1, Fig. 6 B). According to the cell viability assay assessed with LDH release, already at 6 hours after the UV-B exposure, if cis-UCA was not present, then UV-B slightly increased the rupturing of the cell membrane (Fig. 6 C). At 24 and 48 hours after UV-B irradiation, the level of LDH was 2.6 and 2.4 times higher when compared with untreated control cells, respectively (Fig. 6 C). Consequently, cis-UCA resulted in 1.2 ($P < 0.05$) and 1.2 ($P < 0.001$) times lower LDH release when this was assessed at 24 and 48 hours after cis-UCA treatment, respectively (Fig. 6 C). Interestingly, even at the 0.5 hour time point, treatment with cis-UCA increased cell viability as compared with the untreated control (untreated control: 1 vs. UV-B + cis-UCA: 0.53, $P < 0.001$; Fig. 6 C).

DISCUSSION

NLRP3 inflammasomes are intracellular protein complexes, and their activation plays a key role in immune responses.³² In the present study, UV-B irradiation was found to induce the NLRP3-mediated activation of caspase-1 and concurrently elevated the levels of IL-1 β in the HCE cell line and pHCE cells. This finding is in line with the reports of Feldmeyer et al.³³ and Hasegawa et al.³⁴ that UV-B is able to activate

NLRP3 inflammasomes leading to the release of IL-1 β from the keratinocytes of human skin.^{33,34}

Although both TNF- α and LPS are able to provide a priming signal in UV-B-stressed HCE-2 cells and can increase the secretion of IL-1 β , TNF- α appeared to be a more potent priming agent in this study. In fact, recently it has been claimed that TNF- α is a more selective priming factor for NLRP3 in comparison to LPS.³⁵ Additionally, in our corneal cells, UV-B alone resulted in increased IL-1 β release in nonprimed HCE cells; even greater cytokine production was dependent on priming, which is a characteristic of the NLRP3 inflammasome.³⁶ It has been shown previously that UV-B exposure can trigger the expression of IL-1 β mRNA in non-primed HCE cells,⁶ which suggests that UV-B not only activates but also induces the expression of IL-1 β . It has been reported that UV-B possesses an ability to activate TNF-R1 receptors on the cell membrane,³⁷ explaining the UV-B-dependent expression of pro-IL-1 β .

In the present study, intracellular NLRP3 levels were increased in primed pHCE cells but decreased in primed HCE-2 cells after UV-B exposure. Our NLRP3 antibody binds to the N'-terminus of NLRP3 protein (1-150 aa), which is the location of PYD-domain of NLRP3. PYD is a domain that interacts with ASC in the assembly of the inflammasome complex.³⁸ Therefore, the changes observed here seem to be dependent on the amount of PYD domain available, irrespective of whether there is a simultaneous activation of the inflammasome. Although UV-B alone increased the production of IL-1 β , it did not affect the levels of intracellular NLRP3

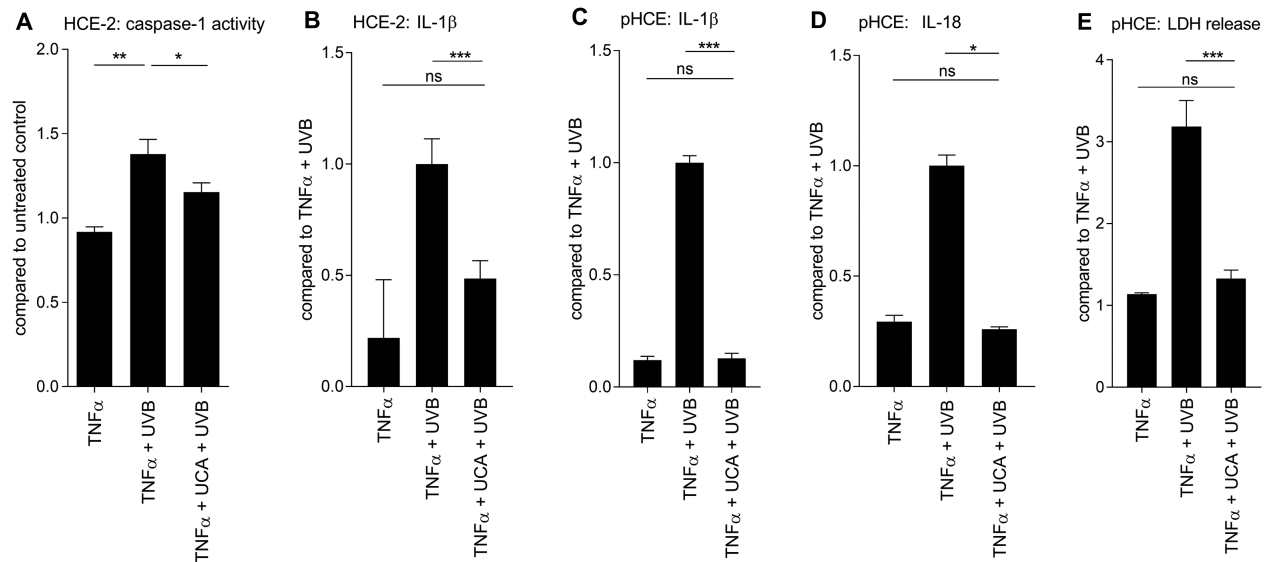


FIGURE 5. Preventive effect of cis-UCA on the secretion of IL-1 β and IL-18 in UV-B-stressed HCE cells. TNF- α -primed (10 ng/mL) HCE cells were pretreated with cis-UCA (100 μ g/mL) for 1 hour followed by irradiation with UV-B (0.2 J/cm²). (A) Caspase-1 activity was measured from HCE-2 cell lysates by an enzymatic assay and (B) IL-1 β levels from HCE-2 cell culture medium samples by the ELISA method at 24 hours after the UV-B exposure. Data for caspase-1 activity were combined from two independent experiments with three parallel samples per group, and data of IL-1 β were combined from three independent experiments with 10 to 12 parallel samples per group in each experiment. The results were confirmed in primary HCE cells where the amounts of (C) IL-1 β , (D) IL-18, and (E) LDH were measured. Levels of IL-1 β (C) and LDH (E) were investigated from four donors with three (TNF- α) or two (UV-B, TNF- α + UV-B, TNF- α + cis-UCA + UV-B) parallel samples from each donor. (D) IL-18 was measured from two donors with two parallel samples from each donor. All data are shown as mean \pm SEM. * P < 0.05, ** P < 0.01, *** P < 0.001, ns = not significant, Mann-Whitney U -test.

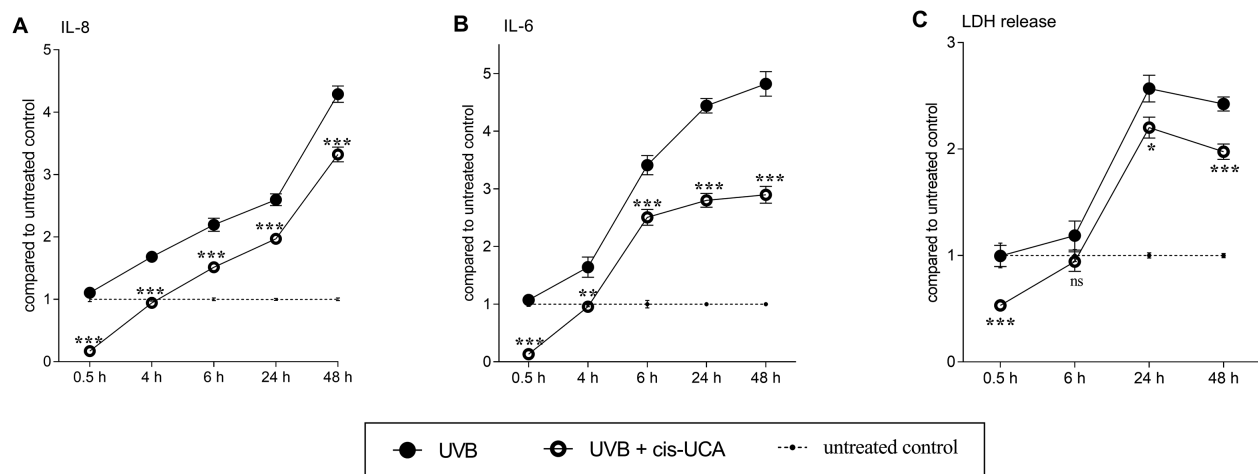


FIGURE 6. Therapeutic effect of cis-UCA on the secretion of IL-8, IL-6, and LDH in UV-B-exposed HCE-2 cells. HCE-2 cells were exposed to UV-B (0.2 J/cm²) and subsequently treated with cis-UCA (100 μ g/mL). Cell culture mediums were collected either 0.5, 4, 6, 24, or 48 hours after the cis-UCA application, and the secretion levels of (A) IL-8 and (B) IL-6 were measured by an ELISA method. (C) Cell membrane integrity was assessed with the LDH assay. (A-C) Results have been converted into the mean of untreated control cells, which is presented in the figure as 1 (dotted line). UV-B (•) and UV-B + cis-UCA (◊) groups were compared at every time point and the differences were analyzed with the Mann-Whitney U -test. Results are represented as mean \pm SEM. * P < 0.05, ** P < 0.01, *** P < 0.001, ns = not significant.

in either cultured pHCE or HCE-2 cells. Previously Benko et al.³⁹ found that UV-B stimulated the expression of NLRP3 mRNA but the protein concentration did not increase after UV-B irradiation in nonprimed HCE cells, which is in line with the present study. Overall, the levels of intracellular NLRP3 do not reveal whether or not inflammasomes have been activated. The activity of caspase-1 is a more accurate; it became activated in both TNF- α -primed HCE-2 and pHCE cells upon UV-B exposure.

In TNF- α -primed HCE cells, the fate of NLRP3 after inflammasome activation depended on the origin of the cells. Thus, UV-B induced higher secretion of NLRP3 in TNF- α primed HCE-2 cells in comparison to pHCE cells. This may be due to different kinetics in the release of inflammasome particles when comparing a cell line to primary cells. We have recently examined ARPE-19 cells and observed that NLRP3 can be either secreted out of the cells or degraded by autophagy following inflammasome

activation.⁴⁰ Additionally, extracellular components of the inflammasome have been detected previously from blood monocyte-derived macrophages, from the serum of patients suffering from cryopyrin-associated periodic syndrome,⁴¹ and in the bronchoalveolar lavage taken from patients with chronic obstructive pulmonary disease and pneumonia.⁴² Moreover, increased NLRP3 protein levels have been detected from the ocular surface epithelium of patients with non-Sjögren syndrome dry eye,⁴³ which could be evidence of the local accumulation of cellular NLRP3 protein in corneal diseases. In the present study, TNF- α alone was also capable of releasing NLRP3 and not simply in pHCE cells followed by UV-B irradiation. It has been reported that TNF- α itself has the capability to induce reactive oxygen species-dependent inflammasome activation and IL-1 β production in neuroblastoma cells,⁴⁴ which could support the idea that NLRP3 is secreted out of the cell upon TNF- α -mediated inflammasome activation. According to our data, it appears that TNF- α triggers the secretion of NLRP3 in the absence of an activating signal. This may be the case with the pHCE cells, where NLRP3 was secreted out of the cell in a TNF- α -independent manner on the levels of intracellular NLRP3 or IL-1 β . We have recently reported similar findings in Hsp90-inhibited ARPE-19 cells where inactive NLRP3 was secreted out of the cells in addition to being degraded by autophagy.⁴⁰

Apart from IL-1 β , the release of another inflammasome-regulated cytokine, IL-18, was studied here from UV-B-stimulated HCE cells. Although priming was needed to enhance the secretion of IL-1 β , pro-forms of IL-18 appeared to be constitutively present in HCE cells. In 2001, Burbach et al. were the first to demonstrate the constitutive expression of IL-18 in primary corneal epithelial cells.¹⁴ Additionally, the same publication revealed that in the corneal epithelial cells, the amounts of secreted and cell-associated IL-18 could be increased by treatment with phorbol 12-myristate 13-acetate, LPS, or double-stranded RNA (poly dI:dC).¹⁴ Because the IL-18 ELISA kit used in our study measures only the mature form of this cytokine, we concluded that UV-B had induced the secretion of bioactive IL-18. Additionally, we found that IL-18 release was not dependent on the caspase-1 activation, which may indicate that IL-18 does not require NLRP3 inflammasome for its cleavage. Previously, it has been shown that several factors (e.g., epithelial cell-derived metalloproteinase Meprin β ,⁴⁵ mast cell derived chymase,⁴⁶ caspase-8,⁴⁷ and NK cell-derived granzyme B⁴⁸) are capable of inducing inflammasome-independent secretion of IL-18. Interestingly, UV-B irradiation has been shown to express granzyme B in human keratinocytes.⁴⁹

Cis-UCA appears to be a promising anti-inflammatory compound because we observed here that cis-UCA reduced the secretion of IL-1 β and IL-18 as well as the activity of caspase-1 in HCE cells. Additionally, we demonstrated that cis-UCA possessed therapeutic properties because it decreased the secretion of IL-6 and IL-8 and alleviated cell death even when administered after the UV-B exposure. We have recently confirmed in a clinical phase I study that 0.5% and 2.5% cis-UCA eye drops are well-tolerated after topical administration to the cornea⁵⁰ and there was no evidence of systemic accumulation or systemic or local side effects upon repeated topical administration of 5% cis-UCA in randomized phase I/IIa dermatological trials.⁵¹ Although the mechanism behind the capacity of cis-UCA to regulate immune responses has been studied widely past 3 decades, it still remains fully unsolved.

In our previous studies, cis-UCA was able to prevent IL-8 and IL-6 secretion via the c-Jun/activator protein-1 and JNK/mitogen activated protein kinase pathways.^{29,30} Instead, Walterscheid et al. have previously reported that the immune suppressive effect of cis-UCA is mediated through the activation of 5-hydroxytryptamine receptor 2 A (5-HTR2A) in mice,⁵² whereas Cloëz-Tayarani and colleagues showed that the stimulation of 5-HT rather increased than decreased the production of IL-1 β through 5-HTR2A in LPS-stimulated human PBMCs.⁵³ In another study, 5-HT stimulation increased the production of IL-1 β but not that of IL-18 in LPS-treated human blood monocytes.⁵⁴ In the study of Dürk et al., IL-1 β secretion was mediated via 5-HTR3, 5-HTR4, and 5-HTR7 instead of 5-HTR2A.⁵⁴ In our present study, productions of IL-1 β and IL-18 were regulated via distinct signaling pathways but cis-UCA reduced the secretion of both of these cytokines, which along with previous findings propose that IL-1 β and IL-18 secretion may not be regulated by 5-HTR2A. Alternatively, cis-UCA has been reported to exert a protodynamic effect in cell cultures, which means that it is capable of transporting protons from a mildly acidic extracellular matrix into the cell.⁵⁵ Because extracellular acidosis is a danger signal for the activation of NLRP3 inflammasome through the K⁺ efflux,⁵⁶ it could be hypothesized that cis-UCA acts to regulate the balance of K⁺ levels between the cytosol and the extracellular space. UV-B is known to trigger K⁺ efflux in human corneal limbal epithelial cells,⁵⁷ and the inhibition of K⁺ efflux is one way to inhibit inflammasome activation and reduce the levels of IL-1 β .^{58,59} Moreover, reactive oxygen species production has been shown to increase the amounts of the pro-form of IL-1 β and to promote its maturation via the NLRP3 inflammasome in hyperosmolarity-induced HCE-2 cells.⁶⁰ That could serve as another way to investigate the silencing mechanism of cis-UCA in UV-B-activated inflammasome signaling. The present study provides new insights to study molecular mechanism of cis-UCA, which has now been associated for the first time with the UV-B-induced activation of inflammasomes in human corneal epithelial cells.

Because cis-UCA can inhibit the secretion of IL-1 β and IL-18 along with its cytoprotective and therapeutic effects, it may well find extensive applications in several pathological conditions (e.g., pterygium, photokeratitis, CDK, and ocular surface squamous neoplasia where UV-B causes cell damage and inflammation).^{3,4}

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References

- Kolozsvári L, Négrádi A, Hopp B, Bor Z. UV absorbance of the human cornea in the 240- to 400-nm range. *Invest Ophthalmol Vis Sci*. 2002;43:2165–2168.
- Ringvold A. Corneal epithelium and UV-protection of the eye. *Acta Ophthalmol Scand*. 1998;76:149–153.
- Delic NC, Lyons JG, Di Girolamo N, Halliday GM. Damaging effects of ultraviolet radiation on the cornea. *Photochem Photobiol*. 2017;93:920–929.
- Serra HM, Holopainen JM, Beuerman R, Kaarniranta K, Suárez MF, Urrets-Zavalía JA. Climatic droplet keratopathy: an old disease in new clothes. *Acta Ophthalmol*. 2015;93:496–504.
- Mallet JD, Rochette PJ. Wavelength-dependent ultraviolet induction of cyclobutane pyrimidine dimers in the human cornea. *Photochem Photobiol Sci*. 2013;12:1310–1318.
- Black AT, Gordon MK, Heck DE, Gallo MA, Laskin DL, Laskin JD. UVB light regulates expression of antioxidants and inflammatory mediators in human corneal epithelial cells. *Biochem Pharmacol*. 2011;81:873–880.
- Cejková J, Stípek S, Crkovská J, et al. UV Rays, the prooxidant/antioxidant imbalance in the cornea and oxidative eye damage. *Physiol Res*. 2004;53:1–10.
- Zheng R, Po I, Mishin V, et al. The generation of 4-hydroxynonenal, an electrophilic lipid peroxidation end product, in rabbit cornea organ cultures treated with UVB light and nitrogen mustard. *Toxicol Appl Pharmacol*. 2013;272:345–355.
- Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity*. 2013;39:1003–1018.
- Keyel PA. How is inflammation initiated? Individual influences of IL-1, IL-18 and HMGB1. *Cytokine*. 2014;69:136–145.
- Malik A, Kanneganti T. Inflammasome activation and assembly at a glance. *J Cell Sci*. 2017;130:3955–3963.
- Gross O, Thomas CJ, Guarda G, Tschopp J. The inflammasome: an integrated view. *Immunol Rev*. 2011;243:136–151.
- Bauernfeind FG, Horvath G, Stutz A, et al. Cutting edge: NF- κ B activating pattern recognition and cytokine receptors license NLRP3 inflammasome activation by regulating NLRP3 expression. *J Immunol*. 2009;183:787–791.
- Burbach GJ, Naik SM, Harten JB, et al. Interleukin-18 expression and modulation in human corneal epithelial cells. *Curr Eye Res*. 2001;23:64–68.
- Novick D, Kim S, Kaplanski G, Dinarello CA. Interleukin-18, more than a Th1 cytokine. *Semin Immunol*. 2013;25:439–448.
- Puren AJ, Fantuzzi G, Dinarello CA. Gene expression, synthesis, and secretion of interleukin 18 and interleukin 1 β are differentially regulated in human blood mononuclear cells and mouse spleen cells. *Proc Natl Acad Sci USA*. 1999;96:2256–2261.
- Shi G, Chen S, Wandu WS, et al. Inflammasomes induced by 7-ketocholesterol and other stimuli in RPE and in bone marrow-derived cells differ markedly in their production of IL-1 β and IL-18. *Invest Ophthalmol Vis Sci*. 2015;56:1658–1664.
- He Y, Hara H, Núñez G. Mechanism and regulation of NLRP3 inflammasome activation. *Trends Biochem Sci*. 2016;41:1012–1021.
- Jo E, Kim JK, Shin D, Sasakawa C. Molecular mechanisms regulating NLRP3 inflammasome activation. *Cell Mol Immunol*. 2016;13:148–159.
- Martinon F, Burns K, Tschopp J. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL- β . *Mol Cell*. 2002;10:417–426.
- Sutterwala FS, Ogura Y, Szczepanik M, et al. Critical role for NALP3/CIA1/cryopyrin in innate and adaptive immunity through its regulation of caspase-1. *Immunity*. 2006;24:317–327.
- Afonina IS, Müller C, Martin SJ, Beyaert R. Proteolytic processing of interleukin-1 family cytokines: variations on a common theme. *Immunity*. 2015;42:991–1004.
- Gibbs NK, Tye J, Norval M. Recent advances in urocanic acid photochemistry, photobiology and photoimmunology. *Photochem Photobiol Sci*. 2008;7:655–667.
- McLoone P, Simics E, Barton A, Norval M, Gibbs NK. An action spectrum for the production of cis-urocanic acid in human skin in vivo. *J Invest Dermatol*. 2005;124:1071–1074.
- Albert E, Walker J, Thiesen A, Churchill T, Madsen K. cis-Urocanic acid attenuates acute dextran sodium sulphate-induced intestinal inflammation. *PLoS ONE*. 2010;5:e13676.
- Konkol Y, Bernoulli J, Streng T, Jääskeläinen K, Laihia J, Leino L. Intravesical treatment with cis-urocanic acid improves bladder function in rat model of acute bladder inflammation. *NeuroUrol Urodyn*. 2016;35:786–791.
- Laihia JK, Taimen P, Kujari H, Leino L. Topical cis-urocanic acid attenuates oedema and erythema in acute and subacute skin inflammation in the mouse. *Br J Dermatol*. 2012;167:506–513.
- Peltonen JM, Pylkkänen L, Jansén CT, et al. Three randomised phase I/IIa trials of 5% cis-urocanic acid emulsion cream in healthy adult subjects and in patients with atopic dermatitis. *Acta Derm Venereol*. 2014;94:415–420.
- Viiri J, Jauhonen HM, Kauppinen A, et al. Cis-urocanic acid suppresses UV-B-induced interleukin-6 and -8 secretion and cytotoxicity in human corneal and conjunctival epithelial cells in vitro. *Mol Vis*. 2009;15:1799–1805.
- Jauhonen H, Kauppinen A, Paimela T, et al. Cis-urocanic acid inhibits SAPK/JNK signaling pathway in UV-B exposed human corneal epithelial cells in vitro. *Mol Vis*. 2011;17:2311–2317.
- Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem*. 1976;72:248–254.
- Strowig T, Henao-Mejia J, Elinav E, Flavell R. Inflammasomes in health and disease. *Nature*. 2012;481:278–286.
- Feldmeyer L, Keller M, Niklaus G, Hohl D, Werner S, Beer H. The inflammasome mediates UVB-induced activation and secretion of interleukin-1 β by keratinocytes. *Curr Biol*. 2007;17:1140–1145.
- Hasegawa T, Nakashima M, Suzuki Y. Nuclear DNA damage-triggered NLRP3 inflammasome activation promotes UVB-induced inflammatory responses in human keratinocytes. *Biochem Biophys Res Commun*. 2016;477:329–335.
- Bauernfeind F, Niepmann S, Knolle PA, Hornung V. Aging-associated TNF production primes inflammasome activation and NLRP3-related metabolic disturbances. *J Immunol*. 2016;197:2900–2908.
- Patel MN, Carroll RG, Galván-Peña S, et al. Inflammasome priming in sterile inflammatory disease. *Trends Mol Med*. 2017;23:165–180.
- Boersma PM, Haarsma LD, Schotanus MP, Ubels JL. TNF-R1 and FADD mediate UVB-induced activation of K⁺ channels in corneal epithelial cells. *Experimental Eye Research*. 2017;154:1–9.
- Bae JY, Park HH. Crystal structure of NALP3 protein pyrin domain (PYD) and its implications in inflammasome assembly. *J Biol Chem*. 2011;286:39528–39536.
- Benko S, Tozser J, Miklossy G, et al. Constitutive and UV-B modulated transcription of Nod-like receptors and their functional partners in human corneal epithelial cells. *Mol Vis*. 2008;14:1575–1583.

40. Piippo N, Korhonen E, Hytti M, et al. Hsp90 inhibition as a means to inhibit activation of the NLRP3 inflammasome. *Sci Rep*. 2018;8:6720.
41. Baroja-Mazo A, Martín-Sánchez F, Gomez AI, et al. The NLRP3 inflammasome is released as a particulate danger signal that amplifies the inflammatory response. *Nat Immunol*. 2014;15:738–748.
42. Franklin BS, Bossaller L, De Nardo D, et al. The adaptor ASC has extracellular and 'prionoid' activities that propagate inflammation. *Nat Immunol*. 2014;15:727–737.
43. Niu L, Zhang S, Wu J, Chen L, Wang Y. Upregulation of NLRP3 inflammasome in the tears and ocular surface of dry eye patients. *PLoS ONE*. 2015;10:e0126277.
44. Álvarez S, Muñoz-Fernández MÁ. TNF- α may mediate inflammasome activation in the absence of bacterial infection in more than one way. *PLoS ONE*. 2013;8:e71477.
45. Banerjee Sanjita, Bond Judith S. Prointerleukin-18 is activated by meprin beta in vitro and in vivo in intestinal inflammation. *The Journal of Biological Chemistry*. 2008;283:31371–31377.
46. Omoto Y, Yamanaka K, Tokime K, et al. Granzyme B is a novel interleukin-18 converting enzyme. *J Dermatol Sci*. 2010;59:129–135.
47. Bossaller L, Chiang P, Schmidt-Lauber C, et al. Cutting edge: FAS (CD95) mediates noncanonical IL-1 β and IL-18 maturation via caspase-8 in an RIP3-independent manner. *J Immunol*. 2012;189:5508–5512.
48. Omoto Y, Tokime K, Yamanaka K, et al. Human mast cell chymase cleaves pro-IL-18 and generates a novel and biologically active IL-18 fragment. *J Immunol*. 2006;177:8315–8319.
49. Hernandez-Pigeon H, Jean C, Charruyer A, et al. Human keratinocytes acquire cellular cytotoxicity under UV-B irradiation. Implication of granzyme B and perforin. *J Biol Chem*. 2006;281:13525–13532.
50. Jauhonen H, Kari E, Pytkäinen L, et al. A randomized phase I clinical study of cis-urocanic acid eye drops in healthy adult subjects. *Acta Ophthalmol*. 2015;93:368–376.
51. Peltonen JM, Pytkäinen L, Jansén CT, et al. Three randomised phase I/IIa trials of 5% cis-urocanic acid emulsion cream in healthy adult subjects and in patients with atopic dermatitis. *Acta Derm Venereol*. 2014;94:415–420.
52. Walterscheid JP, Nghiem DX, Kazimi N, et al. Cis-urocanic acid, a sunlight-induced immunosuppressive factor, activates immune suppression via the 5-HT_{2A} receptor. *Proc Natl Acad Sci USA*. 2006;103:17420–17425.
53. Cloëz-Tayarani I, Petit-Bertron A, Venters HD, Cavaillon J. Differential effect of serotonin on cytokine production in lipopolysaccharide-stimulated human peripheral blood mononuclear cells: involvement of 5-hydroxytryptamine_{2A} receptors. *Int Immunol*. 2003;15:233–240.
54. Dürk T, Panther E, Müller T, et al. 5-Hydroxytryptamine modulates cytokine and chemokine production in LPS-primed human monocytes via stimulation of different 5-HT_R subtypes. *Int Immunol*. 2005;17:599–606.
55. Laihia JK, Kallio JP, Taimen P, Kujari H, Kähäri V, Leino L. Protodynamic intracellular acidification by cis-urocanic acid promotes apoptosis of melanoma cells in vitro and in vivo. *J Invest Dermatol*. 2010;130:2431–2439.
56. Rajamäki K, Nordström T, Nurmi K, et al. Extracellular acidosis is a novel danger signal alerting innate immunity via the NLRP3 inflammasome. *J Biol Chem*. 2013;288:13410–13419.
57. Ubels JL, Van Dyken RE, Louters JR, Schotanus MP, Haarsma LD. Potassium ion fluxes in corneal epithelial cells exposed to UVB. *Experimental Eye Research*. 2011;92:425–431.
58. Muñoz-Planillo R, Kuffa P, Martínez-Colón G, Smith B, Rajendiran T, Núñez G. K⁺ efflux is the common trigger of NLRP3 inflammasome activation by bacterial toxins and particulate matter. *Immunity*. 2013;38:1142–1153.
59. Petrilli V, Papin S, Dostert C, Mayor A, Martinon F, Tschopp J. Activation of the NALP3 inflammasome is triggered by low intracellular potassium concentration. *Cell Death and Differentiation*. 2007;14:1583–1589.
60. Zheng Q, Ren Y, Reinach PS, et al. Reactive oxygen species activated NLRP3 inflammasomes initiate inflammation in hyperosmolarity stressed human corneal epithelial cells and environment-induced dry eye patients. *Exp Eye Res*. 2015;134:133–140.